Total Synthesis of the Phenylpropanoid Glycoside, Acteoside

Toshinari Kawada,* Ryuji Asano, Shiho Hayashida, and Tomoyasu Sakuno

Department of Forestry Science, Faculty of Agriculture, Tottori University, Minami 4-101, Koyama, Tottori 680-0945, Japan

Received April 26, 1999

Introduction

2-(3,4-Dihydroxyphenyl)ethyl-O-α-L-rhamnopyranosyl- $(1 \rightarrow 3)$ -4-*O*-caffeoyl- β -D-glucopyranoside (1) was first extracted from Verbascum sinuatum and named "verbascoside" by M. Scarpati et al.1 in 1963, but they did not provide a complete structural identification. The complete chemical structure of this compound was elucidated by L. Birkofer et al.² in 1968, who also introduced the new name "acteoside". Between these two common names, we prefer "acteoside". Acteoside has also been found in other plant species such as Paulownia tomentosa Steud.,^{3,4} Conandron ramoidioides,⁵ and Clerodendrum myricoides.6

Recently, various bioactivities and pharmaceutical activities of acteoside have been reported. These include antimicrobial activity,⁷ hepatoprotective activity,⁸ sedative effect,⁹ and defense-repair processing in trees.¹⁰ However, the low content of acteoside in each plant species (0.02-0.4%) has limited the further investigation of these activities.

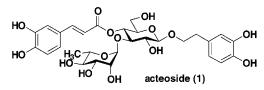
Hence, the chemical synthesis of acteoside has become an important problem. While a total synthesis has not vet been reported, some authors have described the partial synthesis of acteoside^{11,12} or osmanthuside B6,¹³ a close structural relative of acteoside. We report here the first total synthesis of acteoside.

Results and Discussion

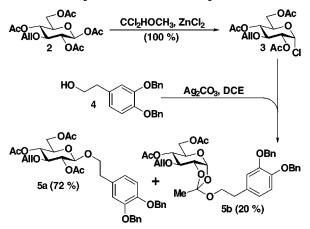
Our strategy for the total synthesis of acteoside (1, Scheme 1) involved a convergent route from the phenethyl glycoside derivative **5a**, from the glucose derivative 3, and the phenetyl derivative 4 (Scheme 2). The 2,4,6-

- (1) Scarpati, M. L.; Monache, D. Ann. Chim. (Rome) 1963, 53, 356. (2) Birkofer, L.; Kaiser, C.; Thomas, U. Z. Naturforsch. 1968, 23b, 1051
- (3) Schilling, G.; Hügel, M.; Mayer, W. Z. Naturforsch. 1982, 37b, 1633.
 - (4) Ota, M.; Taneda, K. Mokuzai Gakkaishi 1989, 35, 438.
 - (5) Nonaka, G.; Nishioka, I. Phytochemistry 1977, 16, 1265.
- (6) Cooper, R.; Solomon, P. H.; Kubo, I.; Nakanishi, K.; Shoolery J. N.; Occolowiz, J. L. J. Am. Chem. Soc. 1980, 102, 7953.
- (7) Shoyama, Y.; Matsumoto, M.; Nishioka, I. Phytochemistry 1987, 26 983
- (8) Xiong, Q.; Hase, K.; Tezuka, Y.; Tani, T.; Namba, T.; Kadota, S. Planta Med. 1998, 64, 120.
- (9) Nakamura, T.; Okuyama, E.; Tsukada, A.; Yamazaki, M.; Satake, M.; Nishibe, S.; Deyama, T.; Moriya, A.; Maruno, M.; Nishimura, H.
- Chem. Pharm. Bull. 1997, 45, 499 (10) Ota, M.; Azuma, T.; Ohira, M.; Abe, K.; Kofujita, H. Mokuzai
- (10) Ota, M., Halman, H., Ohn, M., Hor, K., Hordita, H. Hordzan, Gakkaish 1997, 43, 260. (11) Li, Z. J.; Huang, H. Q.; Cai, M. S. Carbohydr. Res. 1994, 265,
- 227
- (12) Ota, M.; Takahashi, K.; Kofujita, H. J. Wood Sci. 1998, 44, 320.
 (13) Zhang, S. Q.; Li, Z. J.; Wang, A. B.; Cai, M. S.; Feng, R. Carbohydr. Res. 1998, 308, 281.





Scheme 2. Synthesis of Phenethyl Glucoside 5a

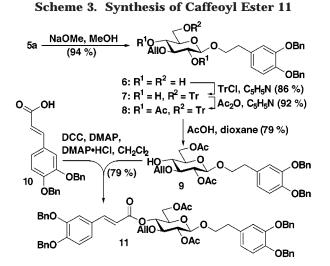


tri-*O*-acetyl-3-*O*-allyl-α-D-glucopyranosyl chloride (3) was prepared in quantitative yield from acetyl 2,4,6-tetra-Oacetyl-3-*O*-allyl- β -D-glucopyranoside¹⁴ (**2**) using α, α dichloromethyl methyl ether and zinc chloride.^{15,16} Condensation of chloride **3** and 3,4-di-O-benzylphenethyl alcohol (4)¹⁷ was achieved by the Koenigs-Knorr method in the presence of silver carbonate. When only 1.2 equiv of silver carbonate was used, a considerable amount of the unexpected ortho ester 5b was produced. The maximum yield of the desired glycoside 5a (72%) was obtained by using 15 equiv of silver carbonate. Under these conditions, formation of the ortho ester 5b was kept at a minimum (20%).

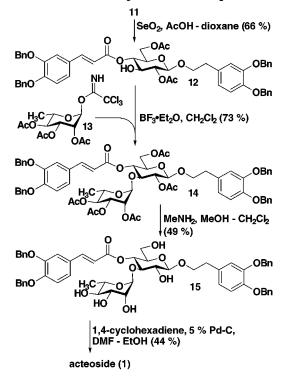
In the next step, the caffeoyl moiety was introduced into the synthesized glycoside 5a, as shown in Scheme 3. Compound 5a was first deacetylated using sodium methoxide to afford the triol 6. To distinguish the 4-OHgroup from the 2-OH- and 6-OH groups in compound 6, the acetyl-migration method was used. Selective tritylation of the 6-OH group of compound 6 gave compound 7 in 82% yield, acetylation of compound 7 gave compound 8 in 92% yield, and treatment of compound 8 with AcOH/ dioxane gave compound 9 (79% yield). Esterification of this glycoside 9 with the caffeoyl derivative 10^{18} was achieved using the Steglich reaction¹⁹ (Keck modification)^{20,21} to give compound **11** (79% yield).

For rhamnosylation (Scheme 4), the 3-O-allyl group of compound 11 was subjected to oxidative cleavage using

- (14) Itoh, T.; Tejima, S. *Chem. Pharm. Bull.* **1983**, *31*, 1631.
 (15) Gross, H. *Z. Chem.* **1978**, *18*, 201.
 (16) Pozsgay, V.; Dubois, E. P.; Pannell, L. *J. Org. Chem.* **1997**, *62*, 2832
- (17) Hamada, A.; Yoden, E. L.; Horng, J. S.; Ruffolo, R. R., Jr.; Patil,
 P. N.; Miller, D. D. J. Med. Chem. 1985, 28, 1265.
 (18) Neish, A. C. Can. J. Biochem. Physiol. 1959, 37, 1431.
- (19) Neises, B.; Steglich, W. Angew. Chem., Int. Ed. Engl. 1978, 17, 522
 - (20) Boden, E. P.; Keck, G. E. J. Org. Chem. 1985, 50, 2394.
 (21) Feldman, K. S.; Smith, R. S. J. Org. Chem. 1996, 61, 2606.



Scheme 4. Rhamnosylation and Deprotections



selenium dioxide.²² Since some cleavage of the caffeoyl ester was observed at a higher reaction temperature (110 °C), the reaction had to be conducted at a lower temperature of around 80 °C to afford compound **12** in acceptable yield (66%). Rhamnosylation of compound **12** was performed with 2,3,4-tri-*O*-acetyl- α -L-rhamnopyranosyl trichloroacetimidate²³ (**13**) in the presence of boron trifluoride diethyl etherate at -20 °C to give the expected α -rhamnoside **14** in 73% yield.

The acetyl groups of compound **14** were then cleaved. Only the acetyl ester needs to be selectively cleaved from caffoeyl ester. In a previous paper on a structurally related phenylpropanoid, osmanthuside B6,¹³ it was stated that the 2-*O*-acetyl group in the glucose residue could not be removed selectively from the caffeoyl ester even under treatment with ammonia–methanol at 5–10 °C. Consequently, various alternative reaction conditions were examined. We finally found that a solution of methylamine in methanol (MeNH₂–MeOH) was the best reagent for achieving the acetyl cleavage. Using MeNH₂–MeOH at -20 °C, all five acetyl groups of compound **14** were successfully removed within 9 h to give compound **15** in 49% yield.

The last step is the debenzylation of compound 15. Neish¹⁸ reported a catalytic hydrogenolysis of a phenolic benzyl ether of an α,β -unsaturated carboxylic acid in conjugation with an aromatic ring. Using these reaction conditions for compound 15, TLC (CHCl₃/MeOH/H₂O, 30: 10:1, v/v/v) analysis indicated that the double bond was stable during the cleavage of three of the four benzyl groups. However, the fourth benzyl group did not undergo hydrogenolysis before the double bond was saturated. Felix *et al.*²⁴ reported catalytic transfer hydrogenation of benzyl ether using 1,4-cyclohexadiene as a proton source. Knapp and Nandan²⁵ found that these reactions conditions can be applied to the hydrogenation reaction in the presence of an easily reduced alkene functionality, although they hydrogenated a benzyl ester, not a benzyl ether. Compound 15 was then treated with 1,4-cyclohexadiene/Pd-C in a solvent mixture of DMF/EtOH at 40 °C. The reaction was monitored by TLC (CHCl₃/ MeOH/H₂O, 30:10:1, v/v/v). At 10 h, the starting compound 15 was completely consumed and the spot corresponding to the desired acteoside (1) ($R_f = 0.20$) had become the major spot, while two additional spots appeared at $R_f = 0.40$ and 0.25. Since the desired spot at $R_f = 0.20$ did not increase thereafter, the reaction was stopped. After isolation, we found that acteoside (1) had been successfully synthesized in 44% yield. ¹H and ¹³C NMR spectra of the synthesized acteoside (1) were identical to those of the natural compound extracted from P. tomentosa Steud.²⁶ and R. glutinosa var. hueichingensis.²⁷ This result indicates that these catalytic hydrogenolysis reactions under the described conditions are suitable for the desired selective debenzylation in the presence of an olefinic double bond.

The advantage of this synthetic strategy is that several structurally related compounds of the acteoside family (phenylpropanoid glycosides) could also be synthesized if saccharides other than the rhamnosyl residue **13** were used. For example, if a glucose derivative or a xylose derivative was employed, plantamajoside²⁸ or conandoroside⁵ could be obtained, respectively.

Experimental Section

All melting points (mp) are uncorrected. NMR spectra were recorded with TMS as an internal standard. The assignments of the signals were determined using a decoupling and/or a 2D-COSY technique. Coupling constants (*J*) are given in Hz. Anhydrous CH_2Cl_2 and $ClCH_2CH_2Cl$ were obtained by distilling from P_2O_5 . Column chromatography was performed on silica gel (Wakogel C-200). Preparative TLC was done on silica gel plates (Kieselgel 60 F₂₅₄, Merck). Unless otherwise indicated, the usual workup for each reaction mixture consists of extraction with

⁽²²⁾ Kariyone, K.; Yazawa, H. *Tetrahedron Lett.* **1970**, *33*, 2885. (23) Van Steijin, A. M. P.; Kamerling, J. P.; Vliegenthart, J. F. G. *Carbohydr. Res.* **1991**, *211*, 261.

⁽²⁴⁾ Felix, A. M.; Heimer, E. P.; Lambros, T. J.; Tzougraki, C.; Meienhofer, J. *J. Org. Chem.* **1978**, *21*, 4194.

 ⁽²⁵⁾ Knapp, S.; Nandan, S. R. J. Org. Chem. 1994, 59, 281.
 (26) Ota, M.; Azuma, T.; Onodera, S.; Taneda, K. Mokuzai Gakkaishi

⁽²⁶⁾ Ota, M.; Azuma, T.; Onodera, S.; Taneda, K. *Mokuzai Gakkaishi* **1993**, *39*, 479.

⁽²⁷⁾ Sasaki, H.; Nishimura, H.; Morota, T.; Chin, M.; Mitsuhashi, H.; Komatsu, Y.; Maruyama, H.; Guo-rui, T.; Wei, H.; Yu-lang, X. *Planta Med.* **1989**, *55*, 458.

⁽²⁸⁾ Ravn, H.; Brimer, L. Phytochemistry 1988, 27, 3433.

EtOAc, washing with brine, drying over Na_2SO_4 , and evaporation *in vacuo*.

2,4,6-Tri-O-acetyl-3-O-allyl-α-D-glucopyranosyl Chloride (3). To a solution of 2^{14} (8.2 g, 21.11 mmol) in CH₂Cl₂ (14 mL) were added α, α -dichloromethyl methyl ether (3.4 mL, 37.59 mmol) and a 2.2 M solution of ZnCl₂·Et₂O (0.96 mL, 2.11 mmol) in CH_2Cl_2 (0.77 mL) at 0 °C. The solution was stirred in the dark at room temperature for 1 h, diluted with EtOAc, neutralized with saturated aqueous NaHCO₃ solution, and worked up to give an oily residue. The residue was purified by column chromatography using a solvent mixture of EtOAc/n-hexane (1: 2, v/v) to give **3** (7.7 g, 100%) as a colorless oil: $[\alpha]^{20}_{D}$ +136.10 (c 1.01, CHCl₃); ¹H NMR (270 MHz, CDCl₃) δ 2.09, 2.10, 2.14 (s, 3 H each), 3.95 (t, J = 9.6, 1 H), 4.05-4.30 (m, 2 H), 4.11 (dd, J = 2.0, 12.4, 1 H), 4.21 (ddd, J = 2.0, 4.2, 9.6, 1 H), 4.25 (dd, J = 4.2, 12.4, 1 H), 4.92 (dd, J = 4.0, 9.6, 1 H), 5.10 (t, J = 9.6, 1 H), 5.17 (ddt, J = 1.3, 1.3, 10.6, 1 H), 5.23 (ddt, J = 1.3, 1.3, 17.1, 1 H), 5.82 (ddt, J = 5.3, 10.6, 17.1, 1 H), 6.29 (d, J = 4.0, 1 H).

3,4-Di-O-benzylphenethyl 2,4,6-Tri-O-acetyl-3-O-allyl-β-D-glucopyranoside (5a). To a solution of 3 (8.8 g, 24.12 mmol) in anhydrous ClCH₂CH₂Cl (62 mL) were added 3,4-di-O-benzylphenethyl alcohol (4)¹⁷ (4.0 g, 11.96 mmol), Ag₂CO₃ (99 g, 359.02 mmol), and powdered molecular sieves (4 Å) at room temperature. The reaction mixture was stirred overnight in the dark at room temperature and filtered, and the filtrate was condensed in vacuo. The residue was diluted with EtOAc, washed with water, and worked up to give an oily residue. Crystallization of the residue from EtOH afforded 5a (5.7 g, 72%) as colorless crystals: mp 68–69 °C; [α]²⁰_D –10.79 (*c* 0.50, CHCl₃); ¹H NMR (270 MHz, CDCl₃) δ 1.90, 2.06, 2.07 (s, 3 H each), 2.77 (t, J=6.8, 2 H), 3.50–3.61 (m, 2 H), 3.54 (t, J=9.5, 1 H), 3.99– 4.06 (m, 3 H), 4.11 (dd, J = 2.7, 12.2, 1 H), 4.21 (dd, J = 4.9, 12.2, 1 H), 4.34 (d, J = 7.8, 1 H), 4.97 (dd, J = 7.8, 9.5, 1 H), 5.04 (t, J = 9.5, 1 H), 5.08-5.23 (m, 2 H), 5.11 (s, 2 H), 5.15 (d, J = 5.7, 2 H), 5.75 (ddt, J = 5.7, 10.3, 17.0, 1 H), 6.69 (dd, J =2.2, 8.4, 1 H), 6.81 (d, J = 2.2, 1 H), 6.84 (d, J = 8.4, 1 H), 7.24-7.49 (m, 10 H). Anal. Calcd for $C_{37}H_{42}O_{11}$ \cdot 0.15 H_2O : C, 67.06; H, 6.39. Found: C, 66.81; H, 6.39.

3,4-Di-O-benzylphenethyl 3-O-Allyl-β-D-glucopyranoside (6). To a solution of 5a (5 g, 7.55 mmol) in MeOH (35 mL) was added dropwise 28% NaOMe in MeOH (2.3 mL) at 0 °C and the mixture stirred at room temperature for 2 h before neutralization with Dowex AG 50W-X8 (Ĥ⁺) resin. The resin was filtered off, the filtrate was concentrated in vacuo, and the residue was recrystallized from Et_2O to give **6** (3.8 g, 94%) as colorless crystals: mp 99–100 °C; [α]²⁰_D +1.85 (*c* 0.50, MeOH); ¹H NMR $(270 \text{ MHz}, \text{CD}_3\text{OD}) \delta 2.74 \text{ (t, } J = 7.3, 2 \text{ H}), 3.08-3.19 \text{ (m, } 2 \text{ H}),$ 3.19-3.30 (m, 2 H), 3.55 (dd, J = 5.4, 11.9, 1 H), 3.60 (m, 1 H), 3.75 (dd, J = 1.9, 11.9, 1 H), 3.94 (dt, J = 7.3, 9.5, 1 H), 4.81 (d, J = 7.3, 1 H), 4.81 (d, J = 7.3,J = 7.3, 1 H), 4.22-4.27 (m, 2 H), 4.96 (s, 2 H), 5.00 (s, 2 H), 5.00 (m, 1 H), 5.20 (ddt, J = 1.6, 1.6, 17.3, 1 H), 5.90 (ddt, J = 5.9, 10.3, 17.3, 1 H), 6.67 (dd, J = 1.9, 8.4, 1 H), 6.81 (d, J = 8.4, 1 H), 6.88 (d, J = 1.9, 1 H), 7.13-7.36 (m, 10 H). Anal. Calcd for C₃₁H₃₆O₈•0.4H₂O: C, 69.39; H, 6.76. Found: C, 69.58; H, 6.86.

3,4-Di-*O*-benzylphenethyl **3**-*O*-Allyl-**6**-*O*-trityl-β-D-glu**copyranoside (7).** To a solution of **6** (3.7 g, 6.9 mmol) in pyridine (41 mL) was added trityl chloride (7.7 g, 27.62 mmol) at 0 °C. The solution was stirred overnight at room temperature in the dark. The reaction mixture was evaporated in vacuo and worked up to give an oily residue, which was purified by column chromatography using a solvent mixture of EtOAc/n-hexane (1:2 to 1:1, v/v) to give 7 (4.6 g, 86%) as a colorless solid: $[\alpha]^{20}D - 22.6$ $(c \ 0.50, \ CHCl_3)$; ¹H NMR (270 MHz, CDCl₃) δ 2.19 (d, J = 1.9, 1 H), 2.67 (d, J = 1.9, 1 H), 2.87 (t, J = 7.0, 2 H), 2.37 (t, J =8.9, 1 H), 3.32-3.46 (m, 4 H), 3.59 (dt, J = 1.9, 8.9, 1 H), 3.63-3.73 (m, 1 H), 4.07 (dt, J = 7.0, 9.7, 1 H), 4.24 (d, J = 7.6, 1 H),4.28 (m, 1 H), 4.39 (m, 1 H), 5.05, 5.11 (s, 2 H each), 5.17 (m, 1 H), 5.29 (m, 1 H), 5.96 (ddt, J = 5.7, 10.5, 17.0, 1 H), 6.72 (dd, J = 2.2, 8.4, 1 H), 6.81 (d, J = 2.2, 1 H), 6.85 (d, J = 8.4, 1 H), 7.17-7.46 (m, 25 H). Anal. Calcd for C₅₀H₅₀O₈•0.4H₂O: C, 77.10; H, 6.47. Found: C, 77.33; H, 6.53.

3,4-Di-*O***-benzylphenethyl 2,4-Di-***O***-acetyl-3-***O***-allyl-6-***O***-trityl-β-D-glucopyranoside (8).** To a solution of **7** (4.6 g, 5.91 mmol) in pyridine (33 mL) was added acetic anhydride (28 mL) at 0 °C. This solution was stirred overnight at room temperature and concentrated to give an oily residue. The residue was suspended in EtOAc, neutralized with saturated aqueous NaH-

CO₃ solution, and worked up to give a colorless solid. Recrystallization of the solid from EtOH gave **8** (4.7 g, 92%) as colorless crystals: mp 143–144 °C; $[\alpha]^{20}_D$ +0.98 (*c* 0.51, CHCl₃); ¹H NMR (270 MHz, CDCl₃) δ 1.78, 1.94 (s, 3 H each), 2.87 (t, *J* = 7.0, 2 H), 3.10 (dd, *J* = 2.4, 10.3, 1 H), 3.16 (dd, *J* = 4.9, 10.3, 1 H), 3.44–3.51 (m, 1 H), 3.50 (t, *J* = 9.5, 1 H), 3.69 (dt, *J* = 7.0, 9.5, 1 H), 4.01 (m, 2 H), 4.12 (dt, *J* = 6.5, 9.5, 1 H), 4.39 (d, *J* = 7.3, 1 H), 4.97–5.08 (m, 4 H), 5.11 (s, 2 H), 5.03–5.20 (m, 2 H), 5.73 (ddt, *J* = 5.7, 10.5, 17.0, 1 H), 6.72 (dd, *J* = 1.9, 8.1, 1 H), 6.82 (d, *J* = 1.9, 1 H), 6.84 (d, *J* = 8.1, 1 H), 7.15–7.47 (m, 25 H). Anal. Calcd for C₅₄H₅₄O₈: C, 75.15; H, 6.31. Found: C, 75.15; H, 6.26.

3,4-Di-O-benzylphenethyl 2,6-Di-O-acetyl-3-O-allyl-β-Dglucopyranoside (9). To a stirred solution of 8 (4.6 g, 5.33 mmol) in 1,4-dioxane (47 mL) was added dropwise AcOH (93 mL). The reaction mixture was heated to 110 °C and stirred at this temperature for 6 h. After being cooled to room temperature, the reaction mixture was diluted with EtOAc, neutralized with a saturated aqueous NaHCO₃ solution, and worked up to give an oily residue. The residue was purified by column chromatography with elution first with CH₂Cl₂ and then with EtOAc/ *n*-hexane (1:2, v/v) to give **9** (2.6 g, 79%) as a colorless oil: $[\alpha]^{20}$ _D -29.29 (c 0.45, CHCl₃); ¹H NMR (270 MHz, CDCl₃) δ 1.90, 2.09 (s, 3 H each), 2.76 (t, J = 6.8, 2 H), 2.79 (d, J = 3.2 1 H), 3.38 (t, J = 9.2, 1 H), 3.42 (ddd, J = 2.2, 4.3, 9.2, 1 H), 3.48-3.57 (m, 1H), 3.54 (m, 1 H), 4.04 (dt, J = 6.8, 9.2, 1 H), 4.17 (m, 2 H), 4.29 (dd, J = 2.2, 11.9, 1 H), 4.32 (d, J = 7.8, 1 H), 4.45 (dd, J= 4.3, 11.9, 1 H), 4.89 (dd, J = 7.8, 9.2, 1 H), 5.11 (s, 2 H), 5.15 (d, J = 5.7, 2 H), 5.16 (ddt, J = 1.6, 1.6, 10.3, 1 H), 5.24 (ddt, J= 1.6, 1.6, 17.0, 1 H), 5.85 (ddt, J = 5.7, 10.3, 17.0, 1 H), 6.69 (dd, J = 1.9, 8.4, 1 H), 6.81 (d, J = 1.9, 1 H), 6.84 (d, J = 8.4, 1H), 7.25-7.48 (m, 10 H). Anal. Calcd for C₃₅H₄₀O₁₀: C, 67.73; H, 6.50. Found: C, 67.45; H 6.53.

3,4-Di-O-benzylphenethyl 2,6-Di-O-acetyl-3-O-allyl-4-O-(3,4-di-O-benzylcaffeoyl)-β-D-glucopyranoside (11). A stirred solution of 9 (2.6 g, 4.19 mmol), 3,4-di-O-benzylcaffeic acid (10)19 (2.2 g, 6.0 mmol), DCC (1.9 g, 9.21 mmol), DMAP (0.25 g, 2.05 mmol), and DMAP·HCl (0.33 g, 2.05 mmol) in CH₂Cl₂ (410 mL) was refluxed overnight. The reaction mixture was cooled to room temperature and filtered through a plug of silica gel, and the solvent was evaporated in vacuo. The residue was purified by column chromatography with elution first with CH₂Cl₂ and then with EtOAc/n-hexane (1:2, v/v) to give a colorless oil. Crystallization of the oil from EtOH gave 11 (3.2 g, 79%) as light yellow crystals: mp 139–140 °C; [α]²⁰_D –34.04 (*c* 0.51, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.91, 2.05 (s, 3 H each), 2.78 (t, J = 7.1, 2H), 3.54-3.61 (m, 1 H), 3.61-3.67 (m, 1 H), 3.63 (t, J = 9.5, 1 H), 4.01–4.09 (m, 3 H), 4.15 (dd, J = 3.2, 12.2, 1 H), 4.19 (dd, J = 4.9, 12.2, 1 H), 4.38 (d, J = 8.0, 1 H), 5.00 (dd, J = 8.0, 9.5, 1 H), 5.12 (s, 2 H), 5.16 (d, J = 8.4, 2 H), 5.18, 5.20 (s, 2 H each), 5.12-5.21 (m, 3 H), 5.71 (ddt, J = 5.6, 10.4, 17.2, 1 H), 6.20 (d, J = 15.9, 1 H), 6.70 (dd, J = 2.0, 8.0, 1 H), 6.82 (d, J = 2.0, 1 H), 6.85 (d, J = 8.0, 1 H), 6.92 (d, J = 8.3, 1 H), 7.07 (dd, J = 2.0, 8.3, 1 H), 7.11 (d, J = 2.0, 1 H), 7.27-7.49 (m, 20 H), 7.59 (d, J = 15.9, 1 H). Anal. Calcd for $C_{58}H_{58}O_{13}$: C, 72.33; H, 6.07. Found: C, 72.07; H, 6.09.

3,4-Di-O-benzylphenethyl 2,6-Di-O-acetyl-4-O-(3,4-di-Obenzylcaffeoyl)- β -D-glucopyranoside (12). To a solution of 11 (2.7 g, 2.8 mmol) in 1,4-dioxane (92 mL) were added SeO₂ (641 mg, 5.78 mmol) and AcOH (253 μ L). The solution was then stirred at 80 °C for 3 h. The reaction mixture was worked up to give an oily residue, which was purified by column chromatography using a solvent mixture of EtOAc/n-hexane (1:2 to 1:1, v/v) to give 12 (1.7 g, 66%) as a light yellow solid: $[\alpha]^{20}{}_D$ –22.30 (c 0.52, CHCl₃); ¹H NMR (270 MHz, CDCl₃) & 1.96, 2.05 (s, 3 H each), 2.66 (d, J = 5.9, 1 H), 2.79 (t, J = 6.6, 2 H), 3.63 (m, 1 H), 3.63–3.74 (m, 1 H), 3.76 (dt, J = 5.9, 9.5, 1 H), 4.06 (dt, J = 6.5, 9.5, 1 H), 4.16-4.23 (m, 2 H), 4.42 (d, J = 7.8, 1 H), 4.88 (dd, J = 7.8, 9.5, 1 H), 5.03 (t, J = 9.5, 1 H), 5.12 (s, 2 H), 5.15 (d, J = 3.5, 2 H), 5.16, 5.19 (s, 2 H each), 6.21 (d, J = 15.9, 1 H), 6.71 (dd, J = 1.9, 8.1, 1 H), 6.82 (d, J = 1.9, 1 H), 6.85 (d, J = 8.1, 1H), 6.91 (d, J = 8.4, 1 H), 7.06 (dd, J = 1.9, 8.4, 1 H), 7.11 (d, J = 1.9, 1 H), 7.25-7.48 (m, 20 H), 7.61 (d, J = 15.9, 1 H). Anal. Calcd for C₅₅H₅₄O₁₃·0.45H₂O: C, 71.55; H, 5.90. Found: C, 71.31; H. 5.83

3,4-Di-O-benzylphenethyl 2,6-Di-O-acetyl-3-O-(2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl)-4-O-(3,4-di-O-benzylcaf-

feoyl)-β-D-glucopyranoside (14). To a stirred solution of 12 (1.7 g, 1.84 mmol) and 2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl trichloroacetimidate (13)²³ (0.96 g, 2.21 mmol) in anhydrous CH_2 -Cl₂ (35 mL) was added BF₃·Et₂O (28 µL, 0.22 mmol) at -20 °C. After 5 h, the reaction mixture was diluted with EtOAc, neutralized with saturated aqueous NaHCO3 solution, and worked up to give an oily residue. The residue was purified using gel-permeation chromatography [Sephadex LH-20, CHCl₃ $\bar{M}e\bar{O}H$ (1:1, v/v)] to give 14 (1.6 g, 73%) as a colorless solid: $[\alpha]^{20}{}_D$ -42.91 (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.99 (d, J =6.4, 3 H), 1.81, 1.95, 1.96, 2.07, 2.09 (s, 3 H each), 2.78 (t, J =6.8, 2 H), 3.57 (m, 1 H), 3.60–3.66 (m, 1 H), 3.80 (dq, J = 6.4, 10.0, 1 H), 3.89 (t, J = 9.5, 1 H), 4.04 (dt, J = 6.8, 9.2, 1 H), 4.14 (dd, J = 3.2, 12.2, 1 H), 4.18 (dd, J = 4.6, 12.2, 1 H), 4.37 (d, J= 8.0, 1 H), 4.86 (d, J = 1.7, 1 H), 4.94 (t, J = 10.0, 1 H), 5.01-5.03 (m, 1H), 5.07 (dd, J = 8.0, 9.5, 1 H), 5.09–5.11 (m, 1H), 5.12, 5.17, 5.20 (s, 2 H each), 5.17 (d, J = 8.8, 2 H), 5.21 (t, J =9.5, 1 H), 6.19 (d, J = 15.8, 1 H), 6.70 (dd, J = 1.7, 8.0, 1 H), 6.81 (d, J = 1.7, 1 H), 6.85 (d, J = 8.0, 1 H), 6.91 (d, J = 8.4, 1H), 7.05 (dd, J = 1.7, 8.4, 1 H), 7.09 (d, J = 1.7, 1 H), 7.28-7.50 (m, 20 H), 7.59 (d, J = 15.8, 1 H). Anal. Calcd for C₆₇H₇₀O₂₀-0.4H₂O: C, 67.33; H, 5.90. Found: C, 67.14; H, 5.96.

3,4-Di-O-benzylphenethyl 3-O-(α-L-Rhamnopyranosyl)-4-O-(3,4-di-O-benzylcaffeoyl)-β-D-glucopyranoside (15). To a solution of 14 (700 mg, 0.59 mmol) in CH₂Cl₂ (9 mL) was added 40% MeNH₂ in MeOH (14 mL) at -20 °C. The reaction mixture was stirred at that temperature for 9 h and then concentrated in vacuo. The residue was purified by TLC using a solvent mixture of MeOH/CH2Cl2 (1:19, v/v) to give 15 (289 mg, 49%) as a colorless solid: $[\alpha]^{20}_{D}$ -54.74 (c 1.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃/CD₃OD) δ 1.12 (d, J = 6.3, 3 H), 2.86 (t, J = 7.1, 2H), 3.30 (t, J = 9.5, 1 H), 3.40-3.50 (m, 2 H), 3.51-3.68 (m, 3 H), 3.61 (dd, J = 3.2, 9.5, 1 H), 3.65-3.76 (m, 1 H), 3.84 (t, J = 9.5, 1 H), 3.29 (dd, J = 1.7, 3.2, 1 H), 4.00–4.15 (m, 1 H), 4.32 (d, J = 8.0, 1 H), 4.96 (t, J = 9.5, 1 H), 5.12, 5.14, 5.17 (s, 2H each), 5.19 (br s, 1 H), 5.20 (s, 2 H), 6.26 (d, J = 15.9, 1 H), 6.77 (dd, J = 2.0, 8.1, 1 H), 6.86 - 6.92 (m, 2 H), 6.95 (d, J = 8.4, 1 H),7.10 (dd, J = 2.0, 8.4, 1 H), 7.16 (br s, 1 H), 7.29–7.47 (m, 20 H), 7.62 (d, J = 15.9, 1 H); ¹³C NMR (400 MHz, CDCl₃/CD₃OD) δ 17.9, 35.7, 61.4, 69.0, 69.5, 71.0, 71.1, 71.3, 71.7, 71.8, 71.9, 72.9, 74.8, 74.9, 80.1, 101.5, 103.1, 114.2, 114.5, 114.9, 115.8,

116.5, 122.1, 123.7, 127.5–128.8, 132.2, 136.8, 137.0, 137.5, 146.7, 147.8, 149.1, 151.7, 167.3. Anal. Calcd for $C_{57}H_{60}O_{15}$ - $0.5H_2O\colon$ C, 69.50; H, 6.14. Found: C, 69.26; H, 6.15.

Acteoside (1). A mixture of 15 (110 mg, 0.11 mmol), 5% Pd-C (110 mg), and 1,4-cyclohexadiene (206 µL, 2.2 mmol) in DMF/EtOH (1:1, v/v, 770 μ L) was stirred at 40 °C for 10 h. The catalyst was filtered off, and the filtrate was concentrated in vacuo to give a yellow oily residue. The residue was purified by preparative TLC using a solvent mixture of CHCl₃/MeOH/H₂O (30:8:1, v/v/v) to give acteoside (1) (30.2 mg, 44%) as a pale-yellow solid: [α]²⁰_D -77.41 (*c* 0.31, MeOH); ¹H NMR (400 MHz, CD₃-OD) δ 1.08 (d, J = 6.1, 3 H), 2.75–2.81 (m, 2 H), 3.29 (t, J = 9.5, 1 H), 3.38 (d, J = 7.8, 9.3, 1 H), 3.48–3.64 (m, 4 H), 3.57 (dd, J= 3.2, 9.5, 1 H), 3.72 (m, 1 H), 3.81 (t, J = 9.3, 1 H), 3.91 (dd, J = 1.7, 3.2, 1 H), 4.04 (m, 1 H), 4.37 (d, J = 7.8, 1 H), 4.80–5.00 (m, 1 H), 5.18 (d, J = 1.7, 1 H), 6.27 (d, J = 15.8, 1 H), 6.56 (dd, J = 2.0, 8.0, 1 H), 6.67 (d, J = 8.0, 1 H), 6.69 (d, J = 2.0, 1 H), 6.77 (d, J = 8.1, 1 H), 6.95 (dd, J = 2.0, 8.1, 1 H), 7.05 (d, J =2.0, 1 H), 7.59 (d, J = 15.8, 1 H); ¹³C NMR (400 MHz, CD₃OD) δ 18.4, 36.5, 62.3, 70.4, 70.6, 72.0, 72.3, 73.8, 76.0, 76.2, 81.3, 103.0, 104.2, 114.7, 115.2, 116.3, 116.5, 117.1, 121.3, 123.2, 127.6, 131.4, 144.7, 146.1, 146.8, 148.0, 149.8, 168.3. Anal. Calcd for C₂₉H₃₆O₁₅·2.1H₂O: C, 55.77; H, 5.81. Found: C, 55.37; H, 6.23.

Acknowledgment. We are grateful to Dr. Yukihiro Sugimoto, Arid Land Research Center, Tottori University, for measurements of the 400 MHz NMR and to the management committee of the 270 MHz NMR in the faculty of engineering, Tottori university, for permission to use their spectrometer. We also thank Ms. Maki Kaneda for preparation of the synthetic materials.

Supporting Information Available: Reproductions of ¹H NMR spectra for compounds **1**, **3**, **5a**,**b**, **6**–**9**, **11**, **12**, **14**, and **15** and ¹³C NMR spectra for compounds **1** and **15**. This material is available free of charge via the Internet at http://pubs.acs.org.

JO9906983